Compound	Reduction in Plasma Glucose Level Compared with Control, %	Statistical Significance Value of p	
Phenformin	10	<0.01a	
IIc	1.5	0.05	
IIIc	4.5	<0.05 ^a	
VId	3.0	<0.01°	
VIIc	2.0	0.05	
VIIIc	8.0	<0.01 ^a	
IXc	4.0	<0.01 ^a	
Xb	2.0	0.05	
XIa	4.0	<0.01ª	

^a Statistically significant.

(IV or V, 0.01 mole) and ethyl bromoacetate (0.011 mole) in absolute ethanol (50 ml) was-refluxed with stirring for 4-6 hr, concentrated, and allowed to crystallize. The products obtained were recrystallized from ethanol (Table II).

Synthesis of VIII and IX—A mixture of IV or V (0.01 mole) and ethyl α -bromopropionate (0.011 mole) in absolute ethanol (50 ml) was refluxed with stirring for 6–8 hr, concentrated, and allowed to crystallize. The products obtained were recrystallized from ethanol as colorless needles (Table II).

Synthesis of X and XI—A mixture of IV or V (0.01 mole) and ethyl β -bromopropionate (0.011 mole) in absolute ethanol (50 ml) was refluxed with stirring for 6–8 hr, concentrated, and allowed to crystallize. The products were recrystallized from 95% ethanol as colorless needles (Table III).

In addition to the aromatic protons at δ 7.0–8.1, the PMR spectra of VIc showed a multiplet at δ 0.8–1.7 for the $(CH_2)_3$ protons of the butyl group together with the methylene group of the cyclic ring, a singlet at δ 2.4 characteristic of the methyl group at C-3 of the pyrazole ring, a singlet at δ 1.3 for the CH₃ of the butyl group, and a singlet at δ 6.4 for the proton at C-4 of the pyrazole ring.

In addition to the aromatic protons at δ 6.8–7.9, the PMR spectra of XIc showed a singlet at δ 2.3 for the two methyl groups together with the two methylene groups of the cyclic ring.

DISCUSSION

Biological Testing—Compounds IIc, IIIc, VId, VIIc, VIIIc, IXc, Xb, and XIa were tested for hypoglycemic activity using alloxanized female albino mice weighing 20-25 g. Alloxan, 100 mg/kg of body weight, was injected into the tail vein in saline solution (10 mg/ml). Three days later, the mice were given the test compounds orally in suspension in 1% carboxymethylcellulose sodium at the rate of 0.2 mmole/kg of body weight.

Each day, four mice were used as a control group and one group of four mice was given the standard drug phenformin in a dose of 100 mg (0.4 mmole)/kg of body weight. Up to six groups of four mice received test compounds. Blood samples were taken at 0, 1, and 3 hr.

Blood was collected into 0.04% NaF solution. Glucose was determined by the microcolorimetric copper reduction technique of Haslewood and Strookman (4). Results are expressed as a percentage reduction of plasma glucose level compared to the control value.

Statistical Significance—Statistical significance was assessed using the Student t test. Statistical significance was accepted where the calculated t value exceeded the tabulated t value at the 0.05 level.

From the data presented in Table IV, it is obvious that VIII possesses marked hypoglycemic activity. While IIIc, IXc, and XIa possess only moderate hypoglycemic activity, the potency of these compounds is more than their corresponding thiourea analogs.

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Synthesis of New Polyamine Derivatives for Cancer Chemotherapeutic Studies

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Received July 28, 1980, from the *Midwest Research Institute, Kansas City, MO 64110, the [‡]University of Missouri-Kansas City, Kansas City, MO 64110, and the [§]Mid-America Cancer Center, University of Kansas Medical Center, Kansas City, KS 66103. Accepted for publication January 15, 1981.

Abstract \square Selected homologs, analogs, and acylated derivatives of spermine and spermidine, together with several heterocyclic and aromatic compounds containing a novoldiamine side chain, were prepared and evaluated biologically. Several compounds possessed activity against B-16 melanoma and human epidermoid carcinoma of the nasopharynx.

Keyphrases \square Polyamines—synthesis of derivatives, cancer chemotherapy \square Chemotherapy—synthesis of polyamine derivatives for cancer chemotherapy \square Derivatives—polyamines, synthesis, cancer chemotherapy

The importance of many naturally occurring polyamines, such as putrescine, spermidine, and spermine, to the growth of living cells is well established (1-4). It was reported recently that increased amounts of certain polyamines were found in rapidly proliferating biological systems (*e.g.*, chick embryo cells, rat regenerating liver cells, and wound-healing tissues) (5-10), as well as in neoplastic cells of animals and humans (11-16). The higher polyamine levels are believed to be contributed by ornithine decarboxylase activity in these tissues (17). Elevated polyamine levels also were detected in the urine of cancer patients (18–20). The high excretion level of polyamines declined when the patients were in remission, and chemotherapy treatment with methotrexate, cytosine arabinoside, 5-azacitidine, or fluorouracil resulted in polyamine depletion in tumor cells (21–23).

Since cellular protein synthesis is affected by polyamines at the transcription and translation level (24) and at least one role of polyamines is to organize the structure and activity of tRNA (25), it was postulated that properly designed analogs of polyamines could be useful in oncology studies. This concept was substantiated by previous reports (26–29) that some spermine and spermidine analogs of both synthetic and plant origin demonstrated inter
$$\begin{split} Ia: R_1 &= R_2 = (CH_2)_3 NH_2 \\ Ib: R_1 &= R_2 = (CH_2)_3 N(CH_3)_2 \\ Ic: R_1 &= (CH_2)_3 N(C_2H_5)_2, R_2 = CH(CH_3) - (CH_2)_3 N(C_2H_5)_2 \\ Id: R_1 &= R_2 = CH(CH_3) - (CH_2)_3 N(C_2H_5)_2 \\ Ie: R_1 + R_2 &= (CH_2CH_2)_2 NCH_3 \\ If: R_1 + R_2 &= (CH_2CH_2)_2 CHNC_5H_{10} \end{split}$$

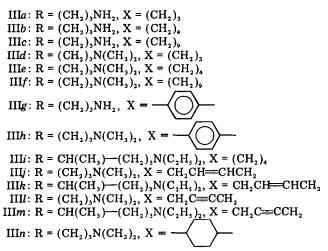
(CH₃)₂CNH(CH₂)₃NH₂

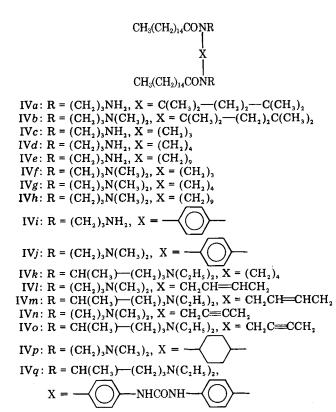
$$(\dot{C}H_2)_2$$

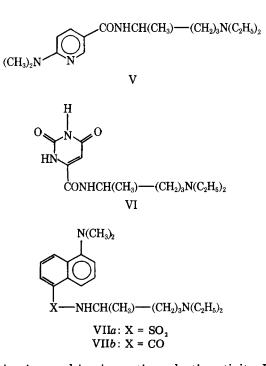
(CH₃)₂ĊNHR

IIa: R = HIIb: $R = (CH_2)_3NH_2$

RNH-X-NHR







esting *in vitro* and *in vivo* antineoplastic activity. Based on the structure-activity relationship information provided by these investigators and the general biochemical and biological aspects of polyamines, selective analogs, homologs, and acylated derivatives of spermine and spermidine (I-IV), as well as several heterocyclic and aromatic compounds containing a novoldiamine side chain (V-VIII), were prepared and their biological activity was evaluated at the National Cancer Institute.

EXPERIMENTAL

Chemistry—N,N-Bis(2-cyanoethyl)amine (VIIIa), prepared by cyanoethylation of ammonia with acrylonitrile (30), was treated with palmitoyl chloride to yield the hexadecaneamide, VIIIb. Catalytic reduction of VIIIb with Raney nickel in ethanolic ammonia (31) gave Ia, isolated as a dihydrochloride salt, mp 240–243° dec. The corresponding dimethyl homolog Ib was obtained from Ia and a mixture of formic acid and formamide, according to the method of Clarke *et al.* (32). Compounds Ic and Id were prepared by reductive alkylation of 5-diethylamino-2pentanone (IX) with 3-diethylaminopropylamine (Xa) and novoldiamine (XI), respectively, followed by acylation. Treatment of 1-methylpiperazine (XII) and 1,4'-bipiperidine (XIII) with palmitoyl chloride in tetrahydrofuran in the presence of triethylamine gave Ie (isolated as a monohydrochloride salt, mp 150–152°) and If (isolated as a free base, mp 68–69°), respectively.

The tetramethyl homologs of spermidine and spermine, IIa (isolated as a diphosphate salt, mp $269-271^{\circ}$) and IIb (disphosphate, mp $239-241^{\circ}$), were prepared by mono- and dicyanoethylation of 2,5-diamino-

 $RN(CH_{2}CH_{2}CN)_{2} CH_{3}CO(CH_{2})_{3}N(C_{2}H_{5})_{2}$ VIIIa: R = H IX $VIIIb: R = CH_{3}(CH_{2})_{14}CO$ $H_{2}N(CH_{2})_{3}NR_{1}R_{2}$ $H_{2}N(CH_{2})_{3}NR_{1}R_{2}$ $H_{2}NCH(CH_{2})_{3}N(C_{2}H_{5})_{2}$ $Xa: R_{1} = R_{2} = C_{2}H_{5}$ XI $Kb: R_{1} = R_{2} = CH_{3}$ $HN - CH_{3}$ $HN - CH_{3}$ KII XIII XIII

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Table I—Antineoplastic Activity Screening Results of Acylated and Some Unacyla	ted Polyamines
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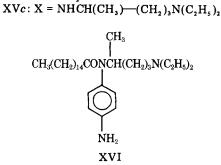
	P-388 Leukemia			B-16 Melanoma	
Compound	Dose, mg/kg	T/C	Dose, mg/kg	T/C	KB Cell Culture ED ₅₀ , µg/ml
Ia	12.5	109			0.96
Īb	6.25	101			0.024
Ic	1.5	98	3.13	104	0.29
Īd	6.25	99	3.13	91	0.026
Ie Ie	25.0	115	0.15		1.7
Ie If	300.0	100		—	100.0
IIa	200.0	100		—	100.0
IIb	3.13	115	<u> </u>		—
IIIe	100.0	110			29.0
IIIi	25.0	92	50.0	Toxic	100.0
			25.0	224	
			12.5	125	
			6.5	133	
			3.13	125	
IIIk	3.13	100	6.25	105	100.0
IIII	100.0	105		_	100.0
IIIm	3.13	108	1.56	105	2.2
IVa	6.25	114	12.5	111	16.0
ĨVĎ	25.0	98			2.3
IVC	25.0	109	—	_	2.3
			-		2.0
IVd	400.0	111	_	—	
IVe	50.0	97			100.0
IVf	1.56	106	1.56	106	2.6
IVg IVh	12.5	90		_	2.3
IVh	3.13	108	6.25	118	12.0
VIi	100.0	98	_		
IVj	3.0	101	_		8.9
IVk	25.0	93	3.13	104	26.0
IVl	6.25	113	50.0	155	2.2
	0.24		25.0	163	212
			12.5	177	
IVm	12.5	102	3.13	98	2.4
IVn	25.0	95	0.10		2.4
IV <i>n</i> IVo	50.0	101	12.5	106	0.63
IVD	3.13	90	12.0	100	
$\frac{1}{1}$		20	_		18.0
IVq	100.0	93	—	—	—
<u>V</u>	25.0	123	—		
VI	50.0	106			
VIIa	6.25	96	6.25	104	
XIVb	100.0	128	_		
XVIIIa	12.5	117		_	<u></u>
XVIIIb	25.0	98			_

$$\begin{array}{c} CH_3 & CH_3 \\ | & | \\ R_1R_2NC(CH_2)_2CNR_1R_2 \\ | & | \\ CH_2 & CH_2 \end{array}$$

XIVa:
$$R_1 = R_2 = H$$

XIVb: $R_1 = H, R_2 = (CH_2)_2 CN$
XIVc: $R_1 = CH_3(CH_2)_1 CO, R_2 = (CH_2)CN$

 $XVa: X = NO_2$ $XVb: X = NH_2$



2,5-dimethylhexane (XIVa) with acrylonitrile, followed by catalytic hydrogenation. Acylation of the intermediate dinitrile XIVb with palmitoyl chloride and subsequent reduction yielded IVa, mp 70–73°. Re-

ductive methylation (32) of IVa afforded IVb. Compounds IVc and IVd were prepared by similar sequential steps through intermediates IIIa–IIIi from the appropriate α,ω -diamines.

The unsaturated spermine analogs were prepared according to a reported method (33). Treatment of trans-1,4-dichloro-2-butene with 3-dimethylaminopropylamine (Xb), followed by acylation of the intermediate diamine III, yielded IVl, mp 58-60°. The novoldiamine analog IVm was prepared from IIIk in an analogous manner. Similarly, the corresponding butyne derivatives IVn (mp 40-41°) and IVo were prepared from 1,4-dichloro-2-butyne through the intermediates IIIl and IIIm, respectively. For the preparation of the cyclohexyl derivative IVp, the intermediate tetramine IIIn was obtained by reductive alkylation



XVIIa: X = OHXVIIb: $X = NHCH(CH_3) - (CH_2)_3N(C_2H_5)_2$



 $\begin{array}{l} \text{XVIII}a: \text{R} = \text{H}, \text{X} = \text{OH} \\ \text{XVIII}b: \text{R} = \text{NO}_2, \text{X} = \text{OH} \\ \text{XVIII}c: \text{R} = \text{NO}_2, \text{X} = \text{NHCH}(\text{CH}_3) & (\text{CH}_2)_3 \text{N}(\text{C}_2\text{H}_5)_2 \\ \text{XVIII}d: \text{R} = \text{NH}_2, \text{X} = \text{NHCH}(\text{CH}_3) & (\text{CH}_2)_3 \text{N}(\text{C}_2\text{H}_5)_2 \end{array}$

of 1,4-cyclohexanedione and Xb. Acylation of IIIn readily yielded IVp, mp 110-115°.

The urea derivative IVq was prepared as follows. *p*-Nitrotrifluoroacetanilide (XVa), prepared from *p*-nitroaniline and trifluoroacetic anhydride, was reduced to the corresponding amino derivative XVb. This reaction was followed by reductive alkylation with IX to give XVc. Acylation of XVc with palmitoyl chloride and subsequent hydrolysis with potassium carbonate gave the *p*-phenylenediamine derivative XVI. Treatment of XVI with diethylcarbamoyl chloride in pyridine, according to a reported method (34), yielded IVq, mp 313-315°.

Condensation of 6-chloronicotinic acid (XVIIa) with novoldiamine in the presence of dicyclohexylcarbodiimide yielded the chloronicotinamide XVIIb. Amination of XVII with aqueous dimethylamine gave V (monophosphate, mp 105–107° dec.). The orotamide derivative VI (mp 196–198°) was obtained by refluxing a mixture of novoldiamine and the methyl ester of orotic acid (35) in 2-methoxyethanol¹. The naphthalenesulfonamide VIIa (dihydrochloride, mp 77–80°) was prepared by treatment of dansyl chloride with novoldiamine. For the preparation of the naphthalenecarboxamide VIIb (monophosphate, mp 211–214° dec.), the starting material 1-naphthoic acid (XVIIIa) was nitrated according to previously reported methods (36, 37). Condensation of the resulting nitroacid XVIIIb and XI, followed by hydrogenation and reductive alkylation, yielded VIIb.

Biological Activity—Test results (Table I) were obtained from the National Cancer Institute. Biological screening was conducted in mice with leukemia P-388 and B-16 melanoma and an *in vitro* system with human epidermoid carcinoma of the nasopharynx (KB cell culture). (For the general procedure and data interpretation, see Refs. 38 and 39.)

None of the polyamines showed good activity against leukemia P-388. On the other hand, N,N'-bis(3-dimethylaminopropyl)-N,N'-bis(palmitoyl)-trans-1,4-diamino-2-butene (IVI) and N,N'-bis(5-diethylamino-2-pentyl)-1,4-butanediamine (IIIi) exhibited inhibitory activity against B-16 melanoma. Compound IVI gave test/control (T/C) values of 155, 163, and 177 at doses of 50, 25, and 12.5 mg/kg, respectively. Compound IIIi had a T/C value of 224 at 25 mg/kg but was toxic at 50 mg/kg.

For the KB cell culture, inhibitory activity was demonstrated when the ED₅₀ value was $<6 \mu g/m$ l. Based on this criterion, several acylated and unacylated polyamines (Ia–Ie, IIIm, IVb, IVc, IVf, IVg, and IVl– IVo) demonstrated inhibitory activity. Three of the most active compounds (Ib–Id) are all monoacylated polyamines that showed ED₅₀ values at concentrations of 0.024, 0.29, and 0.026 $\mu g/m$ l, respectively.

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